

CUPID (Cell-based Un-/identified Protein Interaction Discovery)



Protein-protein Interactions (PPIs) as a Drug Target

- Unlike genes of the genome, there are numerous cellular functions of proteins that are carried out by proteins interacted with other proteins.
- Development of targeting PPI technology is critical in enhancing or shutting down key downstream events of signal transduction pathways and then modulate cellular functions.

Limitations of Conventional Drug Target Approaches

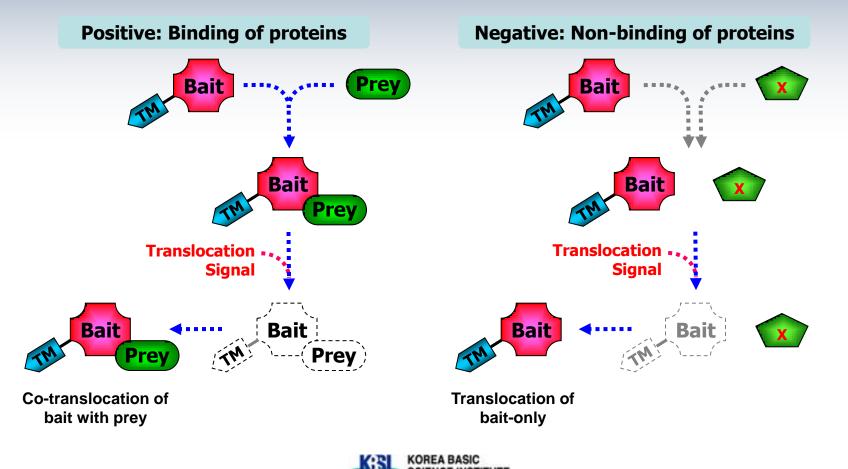
- Difficulty of optimizing PPI-pairs between bait and prey (FRET, BiFC)
- Difficulty of utilizing traditional PPI technologies to identify protein-protein interaction site, peptides targeting PPI sites, and small molecule binding site on/in a PPI
- Difficulty in characterizing involvement of more than two proteins in the complex
- Requirement of antibodies (IP), purification steps (GST-pull down), and other substrates
- Requirement of expensive instruments and technologies such as analytical ultracentrifugation (AUC), surface plasmon resonance (SPR), nuclear magnetic resonance (NMR), X-ray crystallography, and fluorescence correlation spectroscopy (FCS).





Concept of *CUPID* Technology

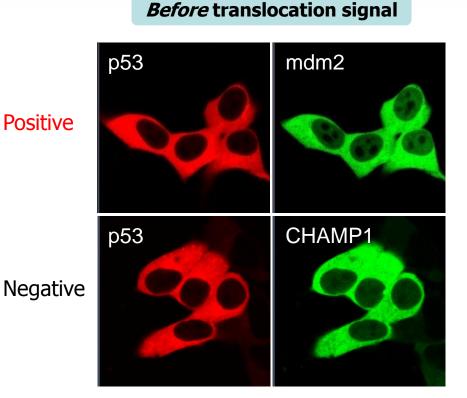
Active co-translocation event of bait and prey reflects the binding status of a bait-prey complex (protein-protein interaction)



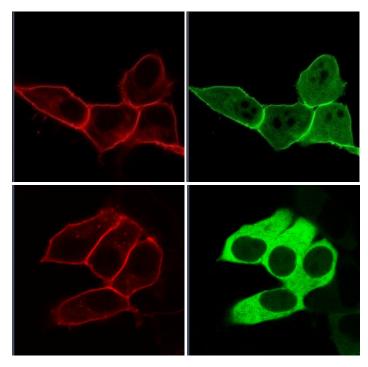


Result Section

- Positive: Active co-translocation of bait (p53, red) and prey (mdm2, green) to cellular membrane reflects the binding status of the complex.
- Negative: Only bait (p53, red) is translocated to cellular membrane while prey (CHAMP1, green) stays in cytoplasm.



After translocation signal





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Advantages of **CUPID**

Native Environment:

- No need for excessive artificial synthetic substrate
- ✓ Intact Cellular Context: Authentic physiological conditions
- ✓ No purification of bait/prey proteins
- \checkmark No need antibodies for bait or prey
- ✓ No expensive equipment for experiment

• Native Conformation of proteins:

- Full-length proteins with correct folding and post-translational modifications
- ✓ Expressed in human cells
- Avoiding denaturation of proteins by artificial buffer
- No modification (truncation, deletion, etc) expression and/or purification of proteins
- No optimization to avoid topological hindrance
- Novel drug-target interactions can be elucidated:
 - Simple detection of protein-protein interactions by conventional fluorescence microscopy
 - ✓ Real-time monitoring possible within a few minutes
 - ✓ Differentiation of different signaling cues
 - ✓ Elimination of false positive results from indirect reporter assays such as IP, Y2H, FRET, BiFC, and FCS



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Applications what we have done with CUPID technology

Dimer bindings

- Cytoplasmic proteins:
 - ✓ Raf1 vs. MEK2, MEK2 vs. ERK2
 - ✓ ERK2 vs. p90RSK, KSR1 vs. 14-3-3
 - ✓ p38AIMP2 vs. complex-forming ARSs
 - ✓ NFkB vs. lkB
- Nuclear proteins:
 - ✓ p53 vs. mdm2, p53 vs. SV40 T antigen
 - ✓ RelA vs. p50

Peptide-Protein bindings

- ✓ p53 domain vs. hdm2
- ✓ TAB1 domain vs. TAK1

(Osteoclast, Bacterial Infection, etc)

Multimer bindings

- 3-mer bindings:
 - ✓ NFkB/IkB (ReIA/p50/IkB) (Multiple Myeloma)
 - ✓ AIMP1/AIMP2/KARS (Lung Cancer, etc)
 - ✓ NICD/RBPJk/MAM (Branin Cancer, etc)
- 4-mer bindings:
 ✓ AIMP1/AIMP2/KARS/DARS
- 5-mer bindings:
 ✓ AIMP1/AIMP2/KARS/DARS/RARS

Inhibitor Screenings

- ✓ Nutlin-3 for p53-hdm2 interaction (Hodgkin Lymphoma, etc)
- ✓ U0126 and PD98059 for ERK2-p90RSK (Breast Cancer, etc)

Compound-mediated Interaction & Its Inhibition

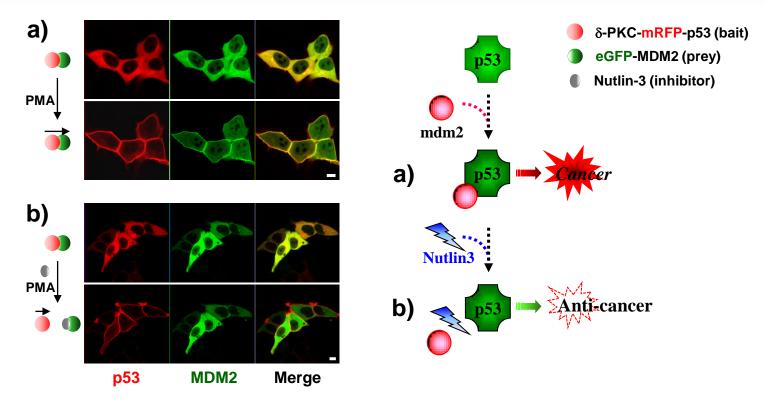
- ✓ Rapamycin-mediated mTOR-FKBP binding (Immunosuppressant, etc)
- ✓ Inhibition of Rapamycin-mediated mTOR-FKBP binding complex by FK506 (Inflammation, etc)





p53-mdm2 binding and their inhibition

The small molecular inhibitor Nutlin-3 is a cis-imidazoline analogue commonly used in anticancer studies that inhibits the interaction between p53 and mdm2

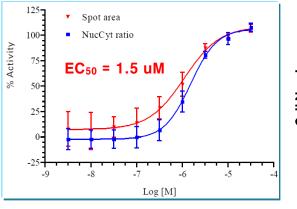


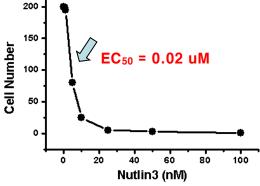




Comparison of Nutlin-3 inhibition curves from 3 different PPI technologies

- In vitro SPR assay : $IC_{50} = 0.09 \ \mu M$
- *In cell* GRIP assay : $EC_{50} = 1.5 \mu M$
 - In cell CUPID assay : $EC_{50} = 0.02 \mu M$
- 100 75 13.6 µM C₅₀: 0.09 uM 50 25 -0 0.01 0.1 10 100 Concentration (µM)





Surface Plasmon Resonance: (Science 303, 844-848, 2004)

GRIP Redistribution assay (www.thermo.com/hcs)

CUPID anlaysis (Angewante Chemie Intl. Ed. 50, 1314-1317, 2011)





Proposed Further Applications

1. Target Identification

 Determination of target for unidentified protein-protein interaction partners or validation of specific protein-protein interaction pairs

2. Inhibitor Screening

 Determination of candidate chemical inhibitors for targeted protein-protein interaction pairs

3. Peptide Inhibitor Screening

 ✓ Determination of core-binding site (binding domain) and generation of peptide inhibitors

4. Drug Repositioning

- ✓ Exploration of unknown targets with known compound
- **5. Complementation & Validation of Results from Conventional Methods**
 - ✓ Confirmation and validation of targets for specific PPI pairs





Validation of CUPID Technology

Patents and Paper

- ✓ Korea, 10-0948767 (Mar. 12, 2010), registered
- ✓ Korea, 10-2010-0037714 (Apr. 23, 2010), filed
- ✓ USA, 12/547,943 (Aug. 25, 2009), filed
- ✓ EU, 09168598.2 (Aug. 25, 2009) , filed
- ✓ Japan, 2009-198750 (Aug. 28, 2009) , filed
- ✓ Angew Chem Int Ed (2011) 50, 1314

National Research Project

- Development of drug target discovery systems using CLSM an FCS (Korea Basic Science Institute, 2009-2010)
- Development of targeted drug screening system
 (The Small & Medium Business Administration, 2009-2011)
- ✓ Development of UTOPIA system for high contents screening of CUPID technology (Korea Basic Science Institute, 2010-2011)
- Pioneer research project (Ministry of Education Science and Technology, PGA043, 2010-2014)

